



Pergamon

SCIENCE @ DIRECT®

Bioorganic & Medicinal Chemistry Letters 13 (2003) 3355–3359

BIOORGANIC &  
MEDICINAL  
CHEMISTRY  
LETTERS

## *N*<sub>1</sub>-Benzenesulfonylgramine and *N*<sub>1</sub>-Benzenesulfonylskatole: Novel 5-HT<sub>6</sub> Receptor Ligand Templates

Manik R. Pullagurta,<sup>a</sup> Małgorzata Dukat,<sup>a</sup> Vincent Setola,<sup>b</sup>  
Bryan Roth<sup>b,c</sup> and Richard A. Glennon<sup>a,\*</sup>

<sup>a</sup>Department of Medicinal Chemistry, School of Pharmacy, Virginia Commonwealth University, Richmond, VA 23298-0540, USA

<sup>b</sup>Department of Biochemistry, School of Medicine, Case Western Reserve University, Cleveland, OH 44106, USA

<sup>c</sup>Departments of Psychiatry and Neurosciences, School of Medicine, Case Western Reserve University, Cleveland, OH 44106, USA

Received 22 January 2003; accepted 25 April 2003

**Abstract**—1-Benzenesulfonyl-5-methoxy-*N,N*-dimethyltryptamine (**3**; *K*<sub>i</sub> = 2.3 nM) is a 5-HT<sub>6</sub> receptor antagonist; removal of the 5-methoxy group (i.e., **6**; *K*<sub>i</sub> = 4.1 nM) has little impact on receptor affinity. In the present study, it is shown that the aminomethyl portion of **6** can be shortened to gramine analogue **10a** (*K*<sub>i</sub> = 3.1 nM); a related skatole derivative **11b** (*K*<sub>i</sub> = 12 nM) also binds with high affinity indicating that the aminoethyl portion of the tryptamines is not required for binding. Compounds **10a** and **11b** represent members of novel classes of 5-HT<sub>6</sub> antagonists.

© 2003 Elsevier Ltd. All rights reserved.

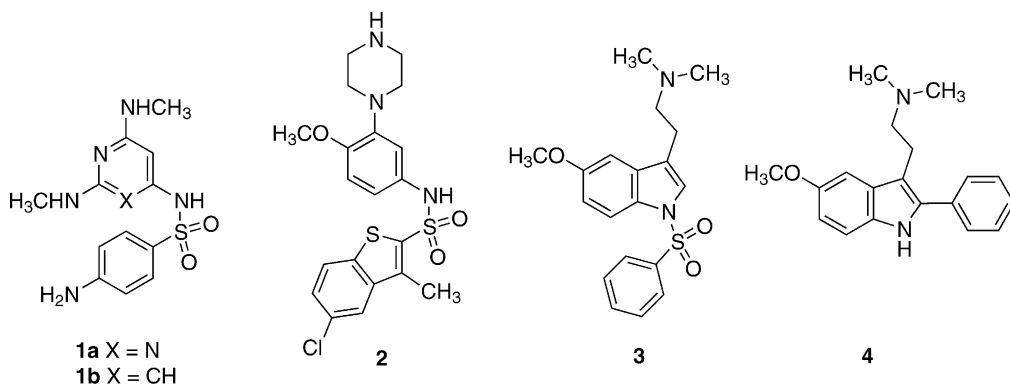
Rat and human 5-HT<sub>6</sub> receptors were first identified and cloned in the early to mid 1990s<sup>1–3</sup> and represent one of seven major families of serotonin receptors (5-HT<sub>1</sub>–5-HT<sub>7</sub>).<sup>4</sup> They are G-protein coupled receptors, positively coupled to an adenylate cyclase second messenger system, and are located primarily in the nucleus accumbens, striatum, hippocampus, and olfactory tubercle of the brain.<sup>5</sup> The exact therapeutic significance of 5-HT<sub>6</sub> receptor ligands is still under investigation, but the binding of various antipsychotics and antidepressants suggests their possible involvement in the treatment of certain mental disorders.<sup>1,3,6</sup> With the development of the first 5-HT<sub>6</sub> antagonists, additional pharmacological actions have been implicated including a role in cognition and convulsive disorders.<sup>7–9</sup> Furthermore, Slassi et al.<sup>8</sup> have suggested that the CNS-specific localization of 5-HT<sub>6</sub> receptors should make them attractive targets for drug development because of the low likelihood that such agents would have peripheral side effects.

The first four types of 5-HT<sub>6</sub> antagonists described in the literature were **1–4**. Compounds **1** (**1a**, Ro 04-6790; **1b**, Ro 63-0563) were reported by Hoffmann-La

Roche,<sup>10</sup> and **2** (SB-271046) by SmithKline Beecham.<sup>11</sup> Compounds **3** (5-methoxy-*N*<sub>1</sub>-benzenesulfonyl-*N,N*-dimethyltryptamine; MS-245) and **4** were reported from our laboratory.<sup>12,13</sup> Interestingly, although representing independent discoveries, three of the four types of initial 5-HT<sub>6</sub> antagonists possess a sulfonamide moiety. Furthermore, **1** and **3** were serendipitous discoveries resulting from random screening, whereas **2** was derived from structure–activity and pharmacokinetic optimization of a lead initially identified from screening of the SmithKline Beecham Compound Data Bank.<sup>11,12,14</sup> Most 5-HT<sub>6</sub> antagonists reported since that time are variations on these themes (e.g., sulfonamides, reversed sulfonamides, 2-substituted tryptamines).<sup>8,9</sup>

Because **1–3** each possesses an *Aryl-N-SO<sub>2</sub>-Aryl'* fragment, it was initially assumed that they might bind at 5-HT<sub>6</sub> receptors in a similar manner.<sup>12</sup> But, with the recent discovery that reversed sulfonamide analogues of **2** retain 5-HT<sub>6</sub> receptor affinity,<sup>15</sup> this concept must now be questioned. Furthermore, although the sulfonamido moiety seems optimal, it has been shown that the aryl-sulfonamido portion of **3** can be replaced by a benzoyl or benzyl group with <10-fold decrease in affinity, and that the –SO<sub>2</sub>– moiety of **3** can even be eliminated—resulting in *N*<sub>1</sub>-(phenyl or substituted phenyl)-tryptamines—with <25-fold reduction in affinity.<sup>16</sup> It now is not clear how **3** binds relative to **1** and **2**. In fact,

\*Corresponding author. Tel.: +1-804-828-8487; fax: +1-804-828-7404; e-mail: glennon@hsc.vcu.edu



Russell et al.<sup>17</sup> have recently proposed an alternative mode of overlap where the tryptamine amino group of **3** is associated with one of the piperazine nitrogen atoms of **2**. The purpose of the present investigation was to examine more closely the terminal amine portion of **3**, and the aminoethyl group in particular, to determine their influence on 5-HT<sub>6</sub> receptor binding.

### Binding Studies

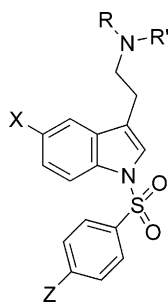
5-HT<sub>6</sub> receptor radioligand binding<sup>18</sup> data are shown in Table 1. Replacement of the 5-methoxy group of **3** ( $K_i$  = 2.3 nM) with hydrogen has little impact on affinity (i.e., **6**;  $K_i$  = 4.1 nM), and *O*-demethylation to the hydroxy analogue **5** ( $K_i$  = 28 nM) decreases affinity by about 10-fold. While this paper was in preparation, Russell et al.<sup>17</sup> reported similar results for **6** and **5**

( $K_i$  = 2.9 and 19 nM, respectively). Apparently, the 5-methoxy group is not a major contributor to binding.

With respect to the terminal amine, the secondary amine **7** ( $K_i$  = 23 nM) binds with 10-fold lower affinity than its parent, **3**. The *N,N*-dimethyl substituents of **3** can be homologated to *N,N*-diethyl (**8**;  $K_i$  = 6.2 nM) with little effect on affinity; however, incorporation of a bulkier benzyl group, as in **9** ( $K_i$  = 43 nM), decreases affinity by about 20-fold.

The alkyl chain separating the terminal amine from the indole nucleus was shortened by a methylene group. Typically, such chain shortening in tryptamine analogues is not well tolerated by serotonin receptors.<sup>19</sup> However, in this instance, the chain-shortened analogue **10a**<sup>20</sup> ( $K_i$  = 3.1 ± 1.2 nM) retains high affinity as compared to its tryptamine counterpart **6**. The obvious

Table 1. Physicochemical and 5-HT<sub>6</sub> receptor binding properties of benzenesulfonyltryptamine analogues<sup>20</sup>



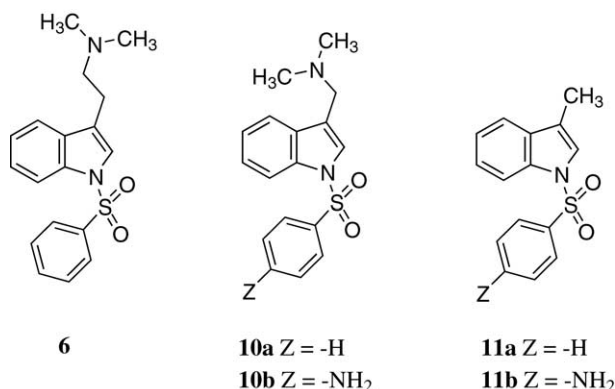
	X	R	R'	Z	Mp (°C)	Recryst. solvent <sup>a</sup>	% Yield	Empirical formula	$K_i$ (nM)	(±SEM)
<b>3<sup>b</sup></b>	–OCH <sub>3</sub>	–CH <sub>3</sub>	–CH <sub>3</sub>	–H	—	—	—	—	2.3	
<b>5</b>	–OH	–CH <sub>3</sub>	–CH <sub>3</sub>	–H	195–197	ME	78	C <sub>18</sub> H <sub>20</sub> N <sub>2</sub> O <sub>3</sub> S.C <sub>2</sub> H <sub>2</sub> O <sub>4</sub>	28	(6)
<b>6</b>	–H	–CH <sub>3</sub>	–CH <sub>3</sub>	–H	194–195	M	22	C <sub>18</sub> H <sub>20</sub> N <sub>2</sub> O <sub>2</sub> S.C <sub>2</sub> H <sub>2</sub> O <sub>4</sub> <sup>c</sup>	4.1	(0.3)
<b>7</b>	–OCH <sub>3</sub>	–CH <sub>3</sub>	–H	–H	215	AM	83	C <sub>18</sub> H <sub>20</sub> N <sub>2</sub> O <sub>3</sub> S.C <sub>2</sub> H <sub>2</sub> O <sub>4</sub>	23	(6)
<b>8</b>	–OCH <sub>3</sub>	–C <sub>2</sub> H <sub>5</sub>	–C <sub>2</sub> H <sub>5</sub>	–H	170	M	54	C <sub>21</sub> H <sub>26</sub> N <sub>2</sub> O <sub>3</sub> S.C <sub>2</sub> H <sub>2</sub> O <sub>4</sub>	6.2	(0.2)
<b>9</b>	–OCH <sub>3</sub>	–CH <sub>3</sub>	–CH <sub>2</sub> Ph	–H	155–156	M	33	C <sub>25</sub> H <sub>26</sub> N <sub>2</sub> O <sub>3</sub> S.C <sub>2</sub> H <sub>2</sub> O <sub>4</sub>	43	(15)
<b>12</b>	–OCH <sub>3</sub>	–CH <sub>3</sub>	–CH <sub>3</sub>	–NHAc	201–203	M	43	C <sub>21</sub> H <sub>25</sub> N <sub>3</sub> O <sub>4</sub> S.C <sub>2</sub> H <sub>2</sub> O <sub>4</sub> <sup>c</sup>	34	(6)
<b>13</b>	–OCH <sub>3</sub>	–CH <sub>3</sub>	–CH <sub>3</sub>	–NH <sub>2</sub>	192–194	P	78	C <sub>19</sub> H <sub>23</sub> N <sub>3</sub> O <sub>3</sub> S.2HCl	2.0	(0.1)
<b>14</b>	–OCH <sub>3</sub>	–C <sub>2</sub> H <sub>5</sub>	–C <sub>2</sub> H <sub>5</sub>	–NHAc	168–169	M	32	C <sub>23</sub> H <sub>29</sub> N <sub>3</sub> O <sub>4</sub> S.C <sub>2</sub> H <sub>2</sub> O <sub>4</sub>	230	(80)
<b>15</b>	–OCH <sub>3</sub>	–C <sub>2</sub> H <sub>5</sub>	–C <sub>2</sub> H <sub>5</sub>	–NH <sub>2</sub>	234–235	M	88	C <sub>21</sub> H <sub>27</sub> N <sub>3</sub> O <sub>3</sub> S.HCl	0.6	(0.2)
<b>16</b>	–H	–CH <sub>3</sub>	–CH <sub>3</sub>	–NHAc	225	AM	33	C <sub>20</sub> H <sub>23</sub> N <sub>3</sub> O <sub>3</sub> S.C <sub>2</sub> H <sub>2</sub> O <sub>4</sub>	27	(11)
<b>17</b>	–H	–CH <sub>3</sub>	–CH <sub>3</sub>	–NH <sub>2</sub>	217–218	AM/E	90	C <sub>18</sub> H <sub>21</sub> N <sub>3</sub> O <sub>3</sub> S.2HCl	0.8	(0.4)
<b>18</b>	–H	–C <sub>2</sub> H <sub>5</sub>	–C <sub>2</sub> H <sub>5</sub>	–NHAc	160–161	M	27	C <sub>22</sub> H <sub>27</sub> N <sub>3</sub> O <sub>3</sub> S.C <sub>2</sub> H <sub>2</sub> O <sub>4</sub> <sup>c</sup>	90	(30)
<b>19</b>	–H	–C <sub>2</sub> H <sub>5</sub>	–C <sub>2</sub> H <sub>5</sub>	–NH <sub>2</sub>	201–203	ME	80	C <sub>20</sub> H <sub>25</sub> N <sub>3</sub> O <sub>2</sub> S.2HCl	0.6	(0.2)
<b>20</b>	–OCH <sub>3</sub>	–CH <sub>3</sub>	–CH <sub>2</sub> Ph	–NH <sub>2</sub>	95–97	ME	20	C <sub>25</sub> H <sub>27</sub> N <sub>3</sub> O <sub>3</sub> S.2HCl	3.0	(1.0)

<sup>a</sup>Recrystallization solvents: M = MeOH, AM = aqueous MeOH; ME = MeOH/anhydrous Et<sub>2</sub>O; P = *n*PrOH.

<sup>b</sup>Binding data for compound **3** was previously reported.<sup>12</sup>

<sup>c</sup>Compounds **6** and **18** crystallized with 0.5 mol H<sub>2</sub>O.

question at this point is whether or not this amine is even required for binding. Removal of the amine of **10a** would result in skatole analogue **11a**, a compound that lacks aqueous solubility.<sup>21</sup> Hence, in order to examine the need for an aminoalkyl group at the indole 3-position, it was necessary to locate a position on the molecule that would tolerate a solubilizing group. An amino group was selected because a water-soluble salt could be formed. It was already known from earlier studies that both electron donating and electron withdrawing substituents are tolerated in the benzenesulfonyl ring of **3**; depending upon the specific substituent, the affinity of the resulting compound is either slightly enhanced or slightly decreased.<sup>12</sup> Consequently, we prepared and examined several aryl amine derivatives of **3**, **6**, and **8** to determine if the amine would be tolerated. The corresponding acetamido analogues were used as controls.



Compound **13** ( $K_i = 2.0$  nM), the 4'-amino analogue of **3**, binds with the same affinity as its parent, **3**. Likewise, the affinities of amino analogues **15**, **17**, and **19** ( $K_i = 0.6$ , 0.8, and 0.6 nM, respectively) are also quite high and indicate that the amino group is tolerated at this position. In compound **20** ( $K_i = 3.0$  nM), the presence of the amino group actually increases affinity by nearly 15-fold relative to its parent, **9**. Furthermore, the affinity of **17** and **19** again indicate that the presence of a 5-methoxy group is not required for binding. The lower affinities of the corresponding acetamido derivatives **12**, **14**, **16**, and **18** show that affinity is not directly related simply to the presence of an NH substituent on the aryl ring. As a further test of the tolerance of the *para* amino substituent, compound **10b**<sup>20</sup> was examined. The affinity of **10b** ( $K_i = 6.9 \pm 0.3$  nM) was similar to that of **10a**.

On the basis that a *para* amino group is tolerated by 5-HT<sub>6</sub> receptors, compound **11b** was prepared and examined. Compound **11b** ( $K_i = 12 \pm 9$  nM) retains affinity for 5-HT<sub>6</sub> receptors. Even though its affinity is about 3-fold lower than tryptamine derivative **6**, it, like **10**, represents a novel type of 5-HT<sub>6</sub> receptor ligand.

### Functional Studies

Benzenesulfonylgramine (1-BSG; **10a**) and the amino benzenesulfonylskatole derivative **11b** (AminoBSK) were examined for functional activity by measuring their ability to either stimulate cAMP accumulation in

HEK cells, or to antagonize 5-HT-induced cAMP production.<sup>22</sup> Both compounds failed to behave as agonists, but both antagonized the actions of 5-HT (data not shown). Calculated IC<sub>50</sub> values for the inhibition of 5-HT-stimulated cAMP production in HEK-5-HT<sub>6</sub> cells by **10a** and **11b** were 880 ( $\pm 35$ ) and 320 ( $\pm 10$ ) nM, respectively. Schild analysis for **11b** provided a  $pA_2$  of 7.0 ( $\pm 0.2$ ). Evidently, both compounds behave as 5-HT<sub>6</sub> antagonists.

### Summary

The present investigation demonstrates that the aminoalkyl portion of *N*<sub>1</sub>-benzenesulfonyltryptamine is, unexpectedly, not a requirement for 5-HT<sub>6</sub> receptor binding. *N*<sub>1</sub>-Benzenesulfonylgramines such as **10a** and **10b** ( $K_i = 3.1$  and 6.9 nM, respectively), derivatives of **3** (more specifically, derivatives of **6**;  $K_i = 4.1$  nM) in which the *N,N*-dimethylaminoethyl portion of the molecule was shortened to an *N,N*-dimethylamino-methyl group, retain affinity. Even more interesting is that the *N,N*-dimethylamine portion of **10b** could be eliminated (**11b**;  $K_i = 12$  nM). Both **10a** and **11b** behaved as antagonists of 5-HT-induced cAMP accumulation. These studies, then, extend the structure–affinity and structure–activity relationships of the *N*<sub>1</sub>-benzenesulfonyltryptamines as 5-HT<sub>6</sub> antagonists and identified two novel classes of 5-HT<sub>6</sub> antagonists: *N*<sub>1</sub>-benzenesulfonylgramines and *N*<sub>1</sub>-benzenesulfonylskatoles. Furthermore, the affinities of these novel analogues question the manner in which they interact at 5-HT<sub>6</sub> receptors relative to **2**; that is, if the amino group of **3**-type compounds is not required for binding, it seems unlikely that this amino group must mimic one of the piperazine nitrogen atoms of **2**. Additional investigations with these classes of compounds are currently underway.

### Acknowledgements

This work was supported in part by NIMH grant MH-60599. Dr. Jagadeesh B. Rangisetty is gratefully acknowledged for his assistance in the preparation of compound **5**.

### References and Notes

1. Monsma, F. J.; Shen, Y.; Ward, R. P.; Hamblin, M. W.; Sibley, D. R. *Mol. Pharmacol.* **1993**, *43*, 320.
2. Ruat, M.; Traiffort, E.; Arrang, J.-M.; Tardivel-Lacombe, J.; Diaz, J.; Leurs, R.; Schwartz, J.-C. *Biochem. Biophys. Acta* **1993**, *193*, 268.
3. Kohen, R.; Metcalf, M. A.; Khan, N.; Druck, T.; Huebner, K.; Lachowicz, J. E.; Meltzer, H. Y.; Sibley, D. R.; Roth, B. L.; Hamblin, M. W. *J. Neurochem.* **1996**, *66*, 47.
4. Hoyer, D.; Hannon, J. P.; Martin, G. R. *Pharmacol. Biochem. Behav.* **2002**, *71*, 533.
5. Ward, R. P.; Hamblin, M. W.; Lachowicz, J. E.; Hoffman, B. J.; Sibley, D. R.; Dorsa, D. M. *Neuroscience* **1995**, *64*, 1105.
6. Roth, B. L.; Craigo, S. C.; Choudhary, M. S.; Uluer, A.; Monsma, F. J.; Shen, Y.; Meltzer, H. Y.; Sibley, D. R. *J. Pharmacol. Exp. Ther.* **1994**, *268*, 1403.

7. Sleight, A. J.; Boess, F. G.; Bourson, A.; Sibley, D. R.; Monsma, F. J. *Drug News Perspect.* **1997**, *10*, 214.

8. Slassi, A.; Isaac, M.; O'Brien, A. *Expert Opin. Ther. Pat.* **2002**, *12*, 513.

9. Russell, M. G. N.; Dias, R. *Curr. Top. Med. Chem.* **2002**, *2*, 643.

10. Sleight, A. J.; Boess, F. G.; Bös, M.; Levet-Trafit, B.; Riemer, C.; Bourson, A. *Br. J. Pharmacol.* **1998**, *124*, 556.

11. Bromidge, S. M.; Brown, A. M.; Clarke, S. E.; Dodgson, K.; Gager, T.; Grassam, H. L.; Jeffrey, P. M.; Joiner, G. F.; King, F. D.; Middlemiss, D. N.; Moss, S. F.; Newman, H.; Riley, G.; Routledge, C.; Wyman, P. *J. Med. Chem.* **1999**, *42*, 202.

12. Tsai, Y.; Dukat, M.; Slassi, A.; MacLean, N.; Demchynshyn, L.; Savage, J. E.; Roth, B. L.; Hufeisen, S.; Lee, M.; Glennon, R. A. *Bioorg. Med. Chem. Lett.* **2000**, *10*, 2295.

13. Glennon, R. A.; Lee, M.; Rangisetty, J. B.; Dukat, M.; Roth, B. L.; Savage, J. E.; McBride, A.; Rauser, L.; Hufeisen, S.; Lee, D. K. H. *J. Med. Chem.* **2000**, *43*, 1011.

14. Bös, M.; Sleight, A. J.; Godel, T.; Martin, J. R.; Riemer, C.; Stadler, H. *Eur. J. Med. Chem.* **2001**, *36*, 165.

15. Bromidge, S. M.; Clarke, S. E.; Gager, T.; Griffith, K.; Jeffrey, P.; Jennings, A. J.; Joiner, G. F.; King, F. D.; Lovell, P. J.; Moss, S. F.; Newman, H.; Riley, G.; Rogers, D.; Routledge, C.; Serafinowska, H.; Smith, D. R. *Bioorg. Med. Chem. Lett.* **2001**, *11*, 55.

16. Lee, M.; Rangisetty, J. B.; Dukat, M.; Slassi, A.; MacLean, N.; Lee, D. K. H.; Glennon, R. A. *Med. Chem. Res.* **2000**, *10*, 230.

17. Russell, M. G.; Baker, R. J.; Barden, L.; Beer, M. S.; Bristow, L.; Broughton, H. B.; Knowles, M.; McAllister, G.; Patel, S.; Castro, J. L. *J. Med. Chem.* **2001**, *44*, 3881.

18. Radioligand binding assay. The h5-HT<sub>6</sub> radioligand binding assays were performed as previously described.<sup>3,6</sup> In brief, h5-HT<sub>6</sub> cDNA was transiently expressed in COS-7 cells using the DEAE-dextran technique.<sup>6</sup> Seventy-two hours after transfection, cells were harvested by scraping and centrifugation from medium containing 10% dialyzed fetal calf serum. Cells were then washed by centrifugation and resuspension once in phosphate buffered saline (pH 7.40; PBS) and then frozen as tight pellets at  $-80^{\circ}\text{C}$  until use. Binding assays were performed at room temperature for 90 min in binding buffer (50 mM Tris-Cl, 10 mM MgCl<sub>2</sub>, 0.1 mM EDTA, pH 7.40) with [<sup>3</sup>H]LSD (1 nM final concentration) using 10  $\mu\text{M}$  clozapine for non-specific binding. Various concentrations of unlabeled test agent (1 to 10,000 nM) were used for  $K_i$  determinations with  $K_i$  values calculated using the LIGAND program.<sup>23</sup> Specific binding represented 80–90% of total binding.  $K_i$  values are the result of triplicate determinations.

19. Glennon, R. A.; Dukat, M. In *Foye's Textbook of Medicinal Chemistry*, Williams, D. A., Lemke, T., Eds.; Williams and Wilkins: Baltimore, 2002; p 315.

20. Synthesis: Melting points (uncorrected) were obtained with a Thomas Hoover apparatus. <sup>1</sup>H NMR spectra were recorded with a Varian EM-390 spectrometer, and peak positions are given in parts per million ( $\delta$ ) downfield from tetramethylsilane as internal standard. Microanalyses were performed by Atlantic Microlab (GA) for the indicated elements, and the results are within 0.4% of theory. Reactions and product mixtures were routinely monitored by thin-layer chromatography on silica gel precoated F<sub>254</sub> Merck plates. The free base of compound **10a** was reported earlier by a different method of synthesis.<sup>21</sup> Compounds in Table 1 were prepared by a procedure similar to that employed for the synthesis of **10b** using the appropriate tryptamine derivative in place of gramine. **1-Benzenesulfonyl-3-(N,N-dimethylamino-methyl)indole oxalate (1-benzenesulfonylgramine; 10a)**. A mixture of gramine (0.50 g, 2.87 mmol) and 60% NaH (0.13 g, 3.16 mmol) was heated at  $130^{\circ}\text{C}$  under N<sub>2</sub> until the evolution

of the H<sub>2</sub> gas ceased. The resultant mass was dissolved in anhydrous DMF (7 mL) and benzenesulfonyl chloride (0.56 g, 3.15 mmol) in anhydrous DMF (3 mL) was added in a dropwise manner at  $0^{\circ}\text{C}$ . The reaction mixture was allowed to stir at room temperature overnight. At  $0^{\circ}\text{C}$  ice was added to the reaction mixture to decompose the excess NaH followed by H<sub>2</sub>O (15 mL); the crude mixture was dissolved in CH<sub>2</sub>Cl<sub>2</sub> (30 mL) and washed with H<sub>2</sub>O (4 $\times$ 50 mL). The organic portion was dried (MgSO<sub>4</sub>) and solvent was removed under reduced pressure. The residue was purified by column chromatography on silica gel (CH<sub>2</sub>Cl<sub>2</sub>/MeOH; 10:1) to give 0.21 g (23%) of an oil. <sup>1</sup>H NMR (CD<sub>3</sub>OD)  $\delta$  2.65 (s, 6H, 2CH<sub>3</sub>), 4.27 (s, 2H, CH<sub>2</sub>), 7.15 (dd,  $J=7.6$  Hz,  $J=7.7$  Hz, 1H, CH), 7.23 (dd,  $J=7.6$  Hz,  $J=7.9$  Hz, 1H, CH), 7.31 (s, 1H, CH), 7.35 (d,  $J=7.9$  Hz, 1H, CH), 7.41–7.47 (m, 1H, CH), 7.55 (d,  $J=7.7$  Hz, 1H, CH), 7.77–7.84 (m, 4H, 4CH). An oxalate salt was prepared in anhydrous MeOH and recrystallized from MeOH/Et<sub>2</sub>O: mp  $198^{\circ}\text{C}$ . Anal. calcd for (C<sub>17</sub>H<sub>18</sub>N<sub>2</sub>O<sub>2</sub>S·C<sub>2</sub>H<sub>2</sub>O<sub>4</sub>) C, H, N. **1-(4-Aminobenzenesulfonyl)-3-(N,N-dimethylamino-methyl)indole hydrochloride (10b)**. A mixture of gramine (1.00 g, 5.73 mmol) and 60% NaH (0.26 g, 6.32 mmol) was heated at reflux in DMF (25 mL) under N<sub>2</sub> for 2 h. *N*-Acetylsulfanilyl chloride (1.60 g, 6.88 mmol) in DMF (5 mL) was added in a dropwise manner at  $0^{\circ}\text{C}$ , and the reaction mixture was allowed to stir at room temperature overnight. The reaction mixture was extracted with CH<sub>2</sub>Cl<sub>2</sub> (2 $\times$ 50 mL); the combined CH<sub>2</sub>Cl<sub>2</sub> fraction was washed with H<sub>2</sub>O (4 $\times$ 50 mL) and the organic portion was dried (MgSO<sub>4</sub>) and solvent was removed under reduced pressure. The residue was purified by column chromatography on silica gel (CH<sub>2</sub>Cl<sub>2</sub>/MeOH; 19:1) to give 1.10 g (52%) of a white solid: mp  $65\text{--}67^{\circ}\text{C}$ . Hydrochloric acid (36%; 5 mL) was added to a solution of the solid acetamide (0.50 g, 1.35 mmol) in absolute EtOH (25 mL) and heated at reflux for 2 h. The reaction mixture was cooled to  $0^{\circ}\text{C}$ , 40% NaOH solution was added to pH 12, and the mixture was extracted with CH<sub>2</sub>Cl<sub>2</sub> (3 $\times$ 50 mL). The combined organic portion was dried (Na<sub>2</sub>SO<sub>4</sub>) and solvent was removed under reduced pressure to give 0.40 g (90%) of **10b** as the free base. <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>)  $\delta$  2.72 (d,  $J=4.2$  Hz, 6H, 2CH<sub>3</sub>), 3.53 (bs, 2H, NH<sub>2</sub>), 4.42 (d,  $J=4.2$  Hz, 2H, CH<sub>2</sub>), 6.55 (d,  $J=8.9$  Hz, 2H, 2CH), 7.31 (dd,  $J=7.5$  Hz,  $J=7.7$  Hz, 1H, CH), 7.38 (dd,  $J=7.5$  Hz,  $J=7.8$  Hz 1H, CH), 7.60 (d,  $J=8.9$  Hz, 2H, 2CH), 7.84 (d,  $J=7.8$  Hz, 1H, CH), 7.88 (d,  $J=7.7$  Hz, 1H, CH), 8.05 (s, 1H, CH). The HCl salt was prepared in anhydrous MeOH and recrystallized from MeOH/Et<sub>2</sub>O: mp  $215\text{--}217^{\circ}\text{C}$ . Anal. calcd for (C<sub>17</sub>H<sub>19</sub>N<sub>3</sub>O<sub>2</sub>S·HCl) C, H, N. **1-(4-Aminobenzenesulfonyl)-3-methylindole hydrochloride (1-(4-amino)benzenesulfonylskatole; 11b)**. A mixture of 3-methylindole (0.50 g, 3.81 mmol) and 60% NaH (0.24 g, 6.00 mmol) was heated at  $110^{\circ}\text{C}$  under N<sub>2</sub> until the evolution of the H<sub>2</sub> gas ceased. At  $0^{\circ}\text{C}$ , anhydrous DMF (7 mL) was added with stirring followed by the dropwise addition of 4-nitrobenzenesulfonyl chloride (0.93 g, 4.19 mmol) in DMF (3 mL). The reaction mixture was allowed to stir at room temperature overnight. Ice and then H<sub>2</sub>O (25 mL) were added at  $0^{\circ}\text{C}$  to decompose excess NaH. The crude product was dissolved in CH<sub>2</sub>Cl<sub>2</sub> (50 mL) and washed with H<sub>2</sub>O (4 $\times$ 50 mL). The organic portion was dried (MgSO<sub>4</sub>) and solvent was removed under reduced pressure. The residue was purified by column chromatography on silica gel (petroleum ether/EtOAc; 19:1) to give 0.22 g (18%) of a solid, mp  $173^{\circ}\text{C}$ , after recrystallization from acetone/petroleum ether. Raney nickel in MeOH ( $\sim 0.8$  g) was added to a methanolic (20 mL) solution of this material (0.20 g, 0.632 mmol) in a Parr bottle, and the reaction mixture was flushed several times with H<sub>2</sub> and then maintained under H<sub>2</sub> at a delivery pressure of 12 psi for 4 h. Catalyst was removed by filtration and the filtrate was concentrated under reduced pressure to give 0.17 g (59%) of a solid. <sup>1</sup>H NMR (CD<sub>3</sub>OD)  $\delta$  2.03 (d,  $J=1.4$  Hz, 3H, CH<sub>3</sub>),

7.00–7.14 (m, 3H, 3CH), 7.09 (d,  $J=8.8$  Hz, 2H, 2CH), 7.19 (d,  $J=1.4$  Hz, 1H, CH), 7.27 (d,  $J=7.0$  Hz, 1H, CH), 7.74 (d,  $J=8.8$  Hz, 2H, 2CH). The HCl salt was prepared in  $\text{CH}_2\text{Cl}_2$ : mp 195–196 °C. Anal. calcd for  $(\text{C}_{15}\text{H}_{14}\text{N}_2\text{O}_2\text{S}\cdot\text{HCl})$  C, H, N.  
21. Hino, T.; Nakamura, T.; Nakagawa, M. *Chem. Pharm. Bull.* **1975**, 23, 2990.

22. cAMP accumulation assay: Increases in cytosolic cAMP levels were measured as previously described in: Roth, B. L.; Baner, K.; Westkaemper, R.; Siebert, D.; Rice, K. C.; Ernsberger, P.; Rothman, R. B. *Proc. Nat. Acad. Sci. U.S.A.* **2002**, 98, 7331.  
23. Munson, P. J.; Rodbard, D. *Analytical Biochem.* **1980**, 107, 220.